

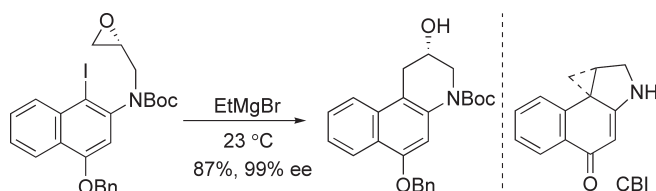
Asymmetric Synthesis of 1,2,9,9a-Tetrahydrocyclopropa[*c*]benzo[*e*]indol-4-one (CBI)

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A short, asymmetric synthesis of the 1,2,9,9a-tetrahydrocyclopropa[*c*]benzo[*e*]indol-4-one (CBI) analogue of the CC-1065 and duocarmycin DNA alkylation subunits is described. Treatment of iodoepoxide **5**, prepared by late-stage alkylation of **4** with (*S*)-glycidal-3-nosylate, with EtMgBr at room temperature directly provides the optically pure alcohol **6** in 87% yield (99% ee) derived from selective metal–halogen exchange and subsequent regioselective intramolecular 6-*endo-tet* cyclization. The use of MeMgBr or *i*-PrMgBr also provides the product in high yields (82–87%), but requires larger amounts of the Grignard reagent to effect metal–halogen exchange and cyclization. Direct transannular spirocyclization of **7** following *O*-debenzylation of **6** provides *N*-Boc-CBI. This approach represents the most efficient (9-steps, 31% overall) and effective (99% ee) route to the optically pure CBI alkylation subunit yet described.

Introduction

CC-1065 (**1**)¹ and duocarmycin SA (**2**)² represent the key members of a class of naturally occurring antitumor agents that also includes duocarmycin A³ and yatakemycin⁴ that derive their biological activity from their ability to selectively alkylate duplex DNA (Figure 1).^{5,6} The study of the natural products, their synthetic unnatural enantiomers,⁷ derivatives, and key analogues has defined the fundamental features that control the DNA alkylation selectivity, efficiency, and catalysis, providing a detailed understanding of the relationships between structure, reactivity, and biological properties.⁶

Among the earliest and most studied of the natural product alkylation subunit analogues is CBI⁸ (Figure 2),

which is not only synthetically more accessible but was found to possess a number of features that have made its use more attractive than the natural alkylation subunits themselves. Most significantly, its derivative analogues were found to be 4-fold more stable and 4-fold more potent than the

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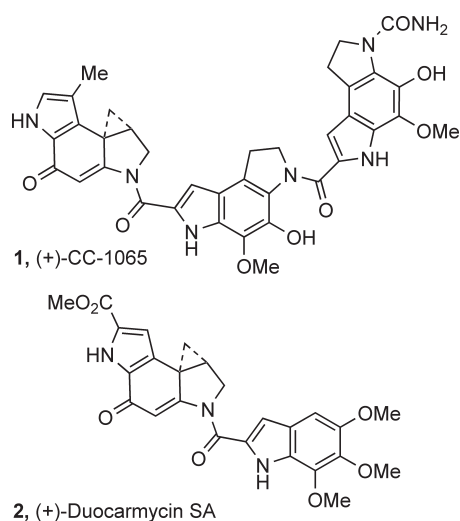


FIGURE 1. Natural products.

corresponding analogues bearing the 7-MeCPI subunit found in CC-1065 and to display a substantially enhanced intrinsic reaction regioselectivity ($>20:1$ vs $4-6:1$).⁹ Additionally, its derivative analogues were found to alkylate DNA with an unaltered sequence selectivity at enhanced rates and with a greater efficiency than the corresponding 7-MeCPI analogues.¹⁰ Although not quite as potent as the corresponding duocarmycin SA analogues, the ability to easily prepare and evaluate CBI analogues designed to probe fundamental questions on structure, reactivity, and activity have resulted in its extensive use.⁶ Thus, it is on this alkylation subunit analogue that new design concepts are often explored, developed, and evaluated.

Its subsequent exploration by others and our continued examination of its properties have provided alternative¹¹ and

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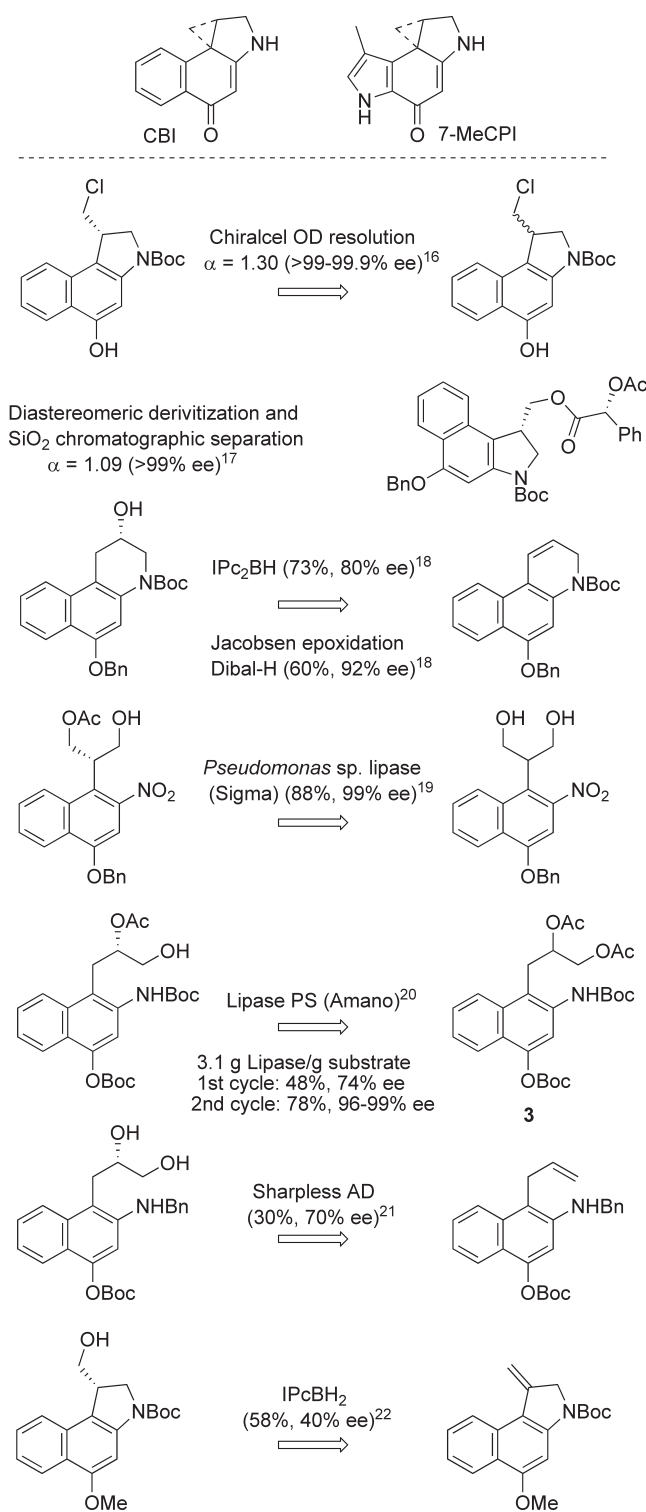


FIGURE 2. Approaches to the asymmetric synthesis of CBI.

iteratively improved syntheses¹²⁻¹⁵ of CBI. Nonetheless, most studies detailing its preparation and use rely on a Chiralcel

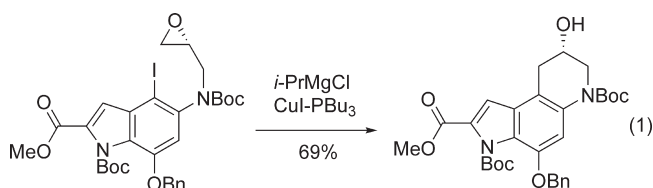
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OD chromatographic resolution¹⁶ of a racemic precursor. Although several approaches to the asymmetric synthesis of CBI have been reported, none have replaced this or related chromatographic resolutions.¹⁷ The asymmetric approaches (Figure 2) include our own strategies relying on an asymmetric hydroboration (73%, 80% ee),¹⁸ a Jacobsen epoxidation (60%, 92% ee),¹⁸ and a lipase-catalyzed resolution of a prochiral diol (88%, 99% ee).¹⁹ Complementary to these efforts are Lown's lipase-catalyzed resolution of **3**,²⁰ the use of a Sharpless AD reaction (30–60%, 70% ee),²¹ and Mohamadi's application of an asymmetric hydroboration reaction (58%, 40% ee).²² The unusual and structure-dependent activity of the unnatural as well as natural enantiomers with this class of natural products requires the evaluation of enantiomers of unusually high optical purities.²³ Thus, even small amounts of natural enantiomer contaminant (<1%) in the unnatural enantiomer samples can provide skewed biological results, requiring the use of a route or technique that delivers the agents in $\geq 99\%$ ee. In the course of our recent development of a unique class of reductively activated CBI-based prodrugs,^{24,25} we have explored and herein report an effective approach to the asymmetric synthesis of CBI.

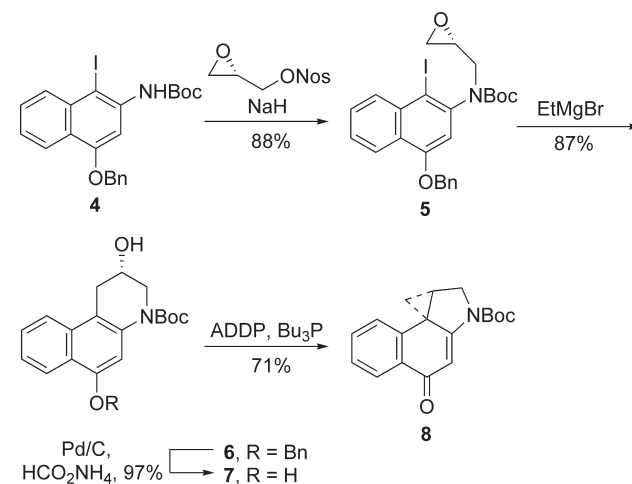
Recently, we described the asymmetric total synthesis of (+)- and *ent*-(-)-yatakemycin and (+)- and *ent*-(-)-duocarmycin SA through use of a selective metal–halogen exchange and subsequent intramolecular regioselective 6-*endo-tet* epoxide opening for the late-stage introduction of the chirality, simplifying access to either enantiomer.²⁶ Thus, treatment of the aryl iodide with *i*-PrMgCl (1.1 equiv of *i*-PrMgCl, -42°C , 1 h) followed by transmetalation to the cuprate (0.2 equiv of CuI–PBU₃, -78°C , 2 h) provided the desired alcohol in 69% yield and 99% ee (eq 1). Herein, we report the extension of this approach to the asymmetric synthesis of the CBI alkylation subunit, representing the most efficient (9-steps, 31% overall) and effective (99% ee) synthesis to date.



Results and Discussion

Synthesis. Alkylation of **4**, prepared in 5-steps from 1,3-dihydroxynaphthalene,¹³ with recrystallized (*S*)-glycidal-3-

SCHEME 1

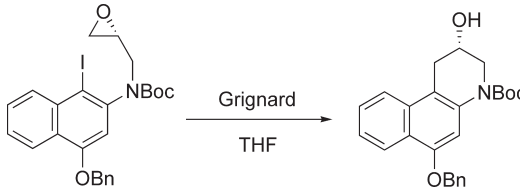


nosylate²⁷ (1.3 equiv, 1.5 equiv of NaH, DMF, 0 to 23°C , 3 h, 90%) provided **5** with clean S_N2 displacement of the nosylate versus epoxide opening and subsequent displacement of the nosylate (Scheme 1). Competitive epoxide opening would provide the enantiomeric epoxide, degrading the enantiomeric purity of the product. However, the product **5** is isolated in nearly perfect optical purity, confirming and establishing the regioselectivity of the alkylation. Initial attempts to afford the aryl Grignard by metal–halogen exchange with *i*-PrMgCl analogous to our efforts on yatakemycin only provided recovered starting material, even at elevated reaction temperatures (23°C) and extended reaction times (72 h), presumably due to the use of a more electron-rich aryl iodide inherent in the naphthalene vs electron-deficient indole substrate. Thus, the more electron-rich naphthalene core slows the rate of metal–halogen exchange even with use of an aryl iodide, rendering most Grignard reagents ineffective. Subsequent efforts to overcome this problem by enlisting lithium–halogen exchange followed by transmetalation onto either a Grignard reagent or a cuprate did provide the desired alcohol **6**. However, the best attainable yields were still low even after optimization (20–40%). These observations led to the examination of a range of more reactive Grignard reagents.²⁸ Whereas such reagents including *i*-PrMgCl·LiCl, *i*-Pr₂Mg·LiCl, and *s*-Bu₂Mg·LiCl provided substantial amounts of side products arising from alkyl or chloride addition to the epoxide, the treatment of **5** with MeMgBr, EtMgBr, or *i*-PrMgBr provided **6** directly in high yields (82–87%), precluding the need for transmetalation onto copper. Formation of the aryl Grignard by metal–halogen exchange (2.0 equiv of EtMgBr, 23°C , 30 min) was followed by the rapid intramolecular ring-opening to give exclusively **6**, the result of intramolecular 6-*endo* versus 5-*exo* addition to the epoxide. MeMgBr and *i*-PrMgBr also provide **6** in high yields (82–87%), albeit requiring larger amounts of the Grignard reagent (10 equiv). Representative studies defining reaction conditions and the number of equivalents of each reagent necessary to optimize yields are summarized in Table 1. Hydrogenolysis removal of the benzyl ether (10% Pd/C, HCO₂NH₄, 9:1 THF/MeOH) provided **7**, and direct transannular Ar-3' spirocyclization upon Mitsunobu activation

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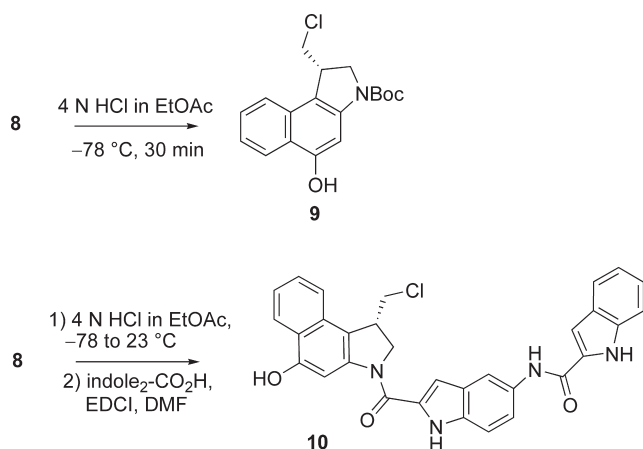
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TABLE 1. Optimization of Grignard Formation and Cyclization



	equiv	time	result
EtMgBr	1.0	120 min	33%
	2.0	30 min	87%
	4.0	30 min	84%
	6.0	30 min	80%
	10.0	10 min	83%
MeMgBr	1.0	24 h	< 1%
	2.0	24 h	7%
	3.0	24 h	30%
	4.0	24 h	38%
	10.0	30 min	87%
<i>i</i> -PrMgBr	1.0	120 min	< 1%
	2.0	120 min	26%
	3.0	120 min	41%
	4.0	120 min	67%
	10.0	10 min	82%

SCHEME 2



of the secondary alcohol (3.0 equiv of 1,1'-(azodicarbonyl)-dipiperidine (ADDP), 3.0 equiv of Bu₃P, toluene, 23 °C, 1 h, 71%) provided *N*-Boc-CBI (**8**), which proved identical in all respects to authentic material. Confirmation of the absolute configuration was made by comparison ($[\alpha]_D$, HPLC Chiralcel OD, t_R) with authentic material for which the stereochemical assignment was previously established by X-ray of a heavy atom derivative.⁹ The optical purity of **8** was determined by chiral phase HPLC (Chiralcel OD column, 7 mL/min, 10% *i*-PrOH/hexane) and established to be 99% ee (see Figure 3, Supporting Information).

Additionally, **8** can be converted to *N*-Boc-*seco*-CBI (**9**), resulting from a stereoelectronically controlled regioselective cyclopropane cleavage, in near quantitative yield by treatment with 4 N HCl in EtOAc at -78 °C for 30 min, followed by solvent removal under a stream of nitrogen while at -78 °C (Scheme 2). A procedure for the direct attachment of the DNA binding subunits onto **8** was also developed. This was accomplished by treatment of **8** with 4 N HCl in EtOAc

at -78 °C for 30 min, followed by warming to room temperature and stirring for 30 min, resulting in regioselective cleavage of the cyclopropane and subsequent *N*-Boc deprotection. Following solvent removal under a stream of nitrogen, the residue was treated with indole₂-CO₂H and EDCI in DMF and stirred for 16 h, providing *seco*-CBI-indole₂ (**10**) in good yield (68%).

Conclusions

An effective (9-step, 31% overall) asymmetric synthesis of *N*-Boc-CBI was developed that provides the optically pure (99% ee) material. The key late stage step introducing the chirality proceeds in excellent yields (82–87%). Thus, treatment of key intermediate **5** with EtMgBr, MeMgBr, or *i*-PrMgBr afforded the desired secondary alcohol **6** directly in superb chemical yield and regioselectivity. This route should prove useful in the future preparation and evaluation of CC-1065 and duocarmycin analogues.

Experimental Section

(*R*)-tert-Butyl (4-(Benzyloxy)-1-iodonaphthalen-2-yl)(oxiran-2-ylmethyl)carbamate (5). A solution of **4** (750 mg, 1.58 mmol) in anhydrous DMF (10 mL) was cooled to 0 °C and treated with NaH (60% dispersion in mineral oil, 76.0 mg, 1.90 mmol). The solution was stirred for 5 min before solid recrystallized (*S*)-glycidal-3-nosylate (99% ee, 491 mg, 1.90 mmol) was added. The reaction mixture was stirred at 0 °C for 30 min, after which it was warmed to room temperature and stirred for 2 h. The solution was quenched with the addition of saturated aqueous NH₄Cl, diluted with EtOAc, washed with H₂O and saturated aqueous NaCl, and dried (Na₂SO₄). The solvent was removed under reduced pressure and the residue was purified by flash chromatography (15% EtOAc/hexane) to provide **5** (735 mg, 88%) as a white solid and as a mixture of rotamers (1:1): ¹H NMR (acetone-*d*₆, 400 MHz) δ 8.34–8.31 (m, 1H), 8.23–8.20 (m, 1H), 7.68 (t, $J = 7.2$ Hz, 1H), 7.64–7.56 (m, 3H), 7.44 (t, $J = 7.2$ Hz, 2H), 7.37 (t, $J = 7.2$ Hz, 1H), 7.23 (s, 0.5H), 7.14 (s, 0.5H), 5.40 (s, 2H), 4.12–4.02 (m, 1H), 3.39–3.25 (m, 2H), 2.69–2.62 (m, 1H), 2.41–2.35 (m, 1H), 1.30 (s, 9H); ¹³C NMR (acetone-*d*₆, 150 MHz) δ 157.13, 157.07, 155.23, 155.16, 145.8, 145.4, 138.70, 138.67, 137.04, 136.98, 134.30, 134.28, 130.50, 130.48, 130.40, 129.84, 129.82, 129.7, 129.6, 129.4, 129.3, 128.3, 128.2, 127.2, 124.22, 124.20, 109.99, 109.96, 96.1, 96.0, 81.6, 81.5, 72.1, 72.0, 54.3, 53.0, 51.3, 50.8, 47.4, 47.0, 29.34, 29.32; IR (film) ν_{\max} 2975, 1700, 1589 cm⁻¹; ESI-TOF HRMS m/z 532.0975 (M + H⁺, C₂₅H₂₆INO₄ requires 532.0979).

5: $[\alpha]_D^{23} +8$ (c 2.0, THF). **ent-5:** $[\alpha]_D^{23} -8$ (c 2.0, THF).

(*S*)-tert-Butyl 6-(Benzyloxy)-2-hydroxy-2,3-dihydrobenzo[*f*]quinoline-4(1*H*)-carboxylate (6). A solution of **5** (126.5 mg, 0.2381 mmol) in distilled anhydrous THF (1.3 mL) at room temperature was treated with EtMgBr (476 μ L, 0.4761 mmol, 1.0 M in THF). The reaction mixture was stirred for 30 min, after which the reaction was quenched with the addition of saturated aqueous NH₄Cl, diluted with EtOAc, washed with H₂O and saturated aqueous NaCl, and dried (Na₂SO₄). The solvent was removed under reduced pressure and the residue was purified by flash chromatography (20% EtOAc/hexane) to provide **6** as a clear oil (84.5 mg, 87%): ¹H NMR (acetone-*d*₆, 400 MHz) δ 8.26 (d, $J = 8.4$ Hz, 1H), 7.87 (d, $J = 8.8$ Hz, 1H), 7.60 (d, $J = 7.2$ Hz, 2H), 7.54 (t, $J = 8.4$ Hz, 1H), 7.44 (t, $J = 7.2$ Hz, 3H), 7.37 (t, $J = 7.2$ Hz, 2H), 5.28 (s, 2H), 4.33 (d, $J = 4.4$ Hz, 1H), 4.28 (m, 1H), 3.99 (dd, $J = 13, 3.2$ Hz, 1H), 3.56 (dd, $J = 12, 8.0$ Hz, 1H), 3.37 (dd, $J = 13, 6.4$ Hz, 1H), 2.92 (dd, $J = 17, 6.0$ Hz, 1H), 1.53 (s, 9H); ¹³C NMR (acetone-*d*₆, 150 MHz) δ 155.7, 153.8, 139.3, 138.2, 134.8, 130.4, 129.7, 129.4, 128.7,

126.0, 125.3, 124.4, 124.0, 115.7, 106.4, 82.2, 71.8, 66.1, 51.9, 35.0, 29.5; IR (film) ν_{\max} 3414, 2975, 2928, 1692, 1622, 1592 cm^{-1} ; ESI-TOF HRMS m/z 406.2014 ($M + H^+$, $C_{25}H_{27}NO_4$ requires 406.2013).

6: $[\alpha]_D +35$ (c 0.41, THF). *ent*-**6:** $[\alpha]_D -34$ (c 0.41, THF). Treatment with MeMgBr (10 equiv) or *i*-PrMgBr (10 equiv) using the above procedure provided **6** in 87 and 82%, respectively.

(S)-tert-Butyl 2,6-Dihydroxy-2,3-dihydrobenzo[*f*]quinoline-4(1H)-carboxylate (7). A solution of **6** (84.7 mg, 0.209 mmol) and ammonium formate (130 mg, 2.09 mmol) in 9:1 distilled THF:MeOH (8 mL) at room temperature was treated with 10% Pd/C (40 mg). The reaction mixture was stirred for 30 min, at which point the solution was filtered through Celite. The solvent was removed under reduced pressure to provide **7** as a white foam (63.9 mg, 97%), identical in all respects to authentic material: ^1H NMR (acetone- d_6 , 400 MHz) δ 8.85 (s, 1H), 8.20 (d, $J = 8.4$ Hz, 1H), 7.84 (d, $J = 8.8$ Hz, 1H), 7.50 (t, $J = 7.6$ Hz, 1H), 7.40 (t, $J = 8.0$ Hz, 1H), 7.27 (s, 1H), 4.25 (m, 2H), 3.98 (dd, $J = 9.2, 2.4$ Hz, 1H), 3.51 (dd, $J = 12, 7.6$ Hz, 1H), 3.35 (dd, $J = 16, 6.0$ Hz, 1H), 2.88 (dd, $J = 16, 6.0$ Hz, 1H), 1.51 (s, 9H); ^{13}C NMR (acetone- d_6 , 150 MHz) δ 155.6, 152.4, 138.3, 135.0, 128.4, 125.3, 124.7, 124.4, 124.2, 114.3, 108.2, 81.9, 66.3, 51.9, 35.0, 29.4; IR (film) ν_{\max} 3306, 2975, 2929, 1670, 1626, 1594 cm^{-1} ; ESI-TOF HRMS m/z 316.1540 ($M + H^+$, $C_{18}H_{21}NO_4$ requires 316.1543).

7: $[\alpha]_D +42$ (c 0.94, THF). *ent*-**7:** $[\alpha]_D -41$ (c 1.13, THF).

1,2,9a-Tetrahydrocyclopropa[*c*]benzo[*e*]indol-4-one (N-Boc-CBI, 8). A solution of **7** (10.6 mg, 0.0336 mmol) in toluene (1 mL) at room temperature was treated with ADPP (42.4 mg, 0.168 mmol) and tributyl phosphine (42 μL , 0.168 mmol). The reaction mixture was stirred for 30 min, at which point the solvent was removed under reduced pressure and the residue was purified by flash chromatography (30% EtOAc/hexane) to provide **8** (7.1 mg, 71%) as a white solid, identical in all respects to authentic material: ^1H NMR (acetone- d_6 , 400 MHz) δ 8.08 (d, $J = 8.0$ Hz, 1H), 7.53 (t, $J = 7.6$ Hz, 1H), 7.39 (t, $J = 8.0$ Hz, 1H), 7.10 (d, $J = 8.0$ Hz, 1H), 6.78 (s, 1H), 4.04 (m, 2H), 3.03 (m, 1H), 1.66 (dd, $J = 7.6, 4.0$ Hz, 1H), 1.54 (s, 9H), 1.49 (t, $J = 4.4$ Hz, 1H); ^{13}C NMR (acetone- d_6 , 150 MHz) δ 186.3, 162.0, 153.3, 142.5, 134.6, 133.4, 127.9, 127.8, 123.7, 109.5, 84.0, 54.8, 34.9, 30.4, 29.2, 25.4; IR (film) ν_{\max} 2975, 2929, 1723, 1626, 1600, 1564 cm^{-1} ; ESI-TOF HRMS m/z 298.1439 ($M + H^+$, $C_{18}H_{19}NO_3$ requires 298.1438).

8: $[\alpha]_D +131$ (c 0.53, THF). *ent*-**8:** $[\alpha]_D -129$ (c 0.60, THF).

(S)-tert-Butyl 1-(Chloromethyl)-5-hydroxy-1H-benzo[*e*]indole-3(2H)-carboxylate (N-Boc-seco-CBI, 9). A sample of **8** (2.5 mg, 0.0084 mmol) at -78 °C was treated with 4 N HCl in EtOAc (0.5 mL) and stirred at -78 °C for 30 min. While keeping the solution cooled, the solvent and HCl gas was removed under a

stream of nitrogen, providing **9** as a light brown solid (2.7 mg, 96%), identical in all respects to authentic material: ^1H NMR (acetone- d_6 , 400 MHz) δ 8.19 (d, $J = 8.0$ Hz, 1H), 7.77 (d, $J = 8.4$ Hz, 1H), 7.73 (br s, 1H), 7.50 (t, $J = 7.6$ Hz, 1H), 7.31 (t, $J = 8.4$ Hz, 1H), 4.20 (dd, $J = 11, 2.4$ Hz, 1H), 4.15 (t, $J = 10$ Hz, 1H), 4.07 (m, 1H), 3.99 (dd, $J = 11, 3.2$ Hz, 1H), 3.66 (dd, $J = 11, 8.8$ Hz, 1H), 1.57 (s, 9H); ^{13}C NMR (acetone- d_6 , 150 MHz) δ 156.4, 153.9, 132.6, 129.2, 127.0, 125.2, 124.2, 124.0, 123.4, 115.6, 100.8, 82.5, 54.6, 48.9, 42.9, 29.6; ESI-TOF HRMS m/z 334.1218 ($M + H^+$, $C_{18}H_{20}ClNO_3$ requires 334.1204).

(S)-N-(2-(1-(Chloromethyl)-5-hydroxy-2,3-dihydro-1H-benzo[*e*]indole-3-carbonyl)-1H-indol-5-yl)-1H-indole-2-carboxamide (seco-CBI-indole, 10). A sample of **8** (13.5 mg, 0.0455 mmol) at -78 °C was treated with 4 N HCl in EtOAc (1.0 mL) and stirred at -78 °C for 30 min. The solution was warmed to 23 °C and stirred for 30 min, at which point the solvent and HCl gas was removed under a stream of nitrogen. The resulting residue was dissolved in anhydrous DMF (1.0 mL) and EDCI (26.1 mg, 0.136 mmol) and indole- CO_2H (16.0 mg, 0.0500 mmol) were added. The resulting reaction mixture was stirred for 18 h, at which point the solution was diluted with EtOAc, washed with 1 M HCl, saturated aqueous NaHCO_3 , saturated aqueous NaCl, and dried (Na_2SO_4). The solvent was removed under reduced pressure and the residue was purified by flash chromatography (40% THF/hexane) to provide **10** as a pale yellow solid (16.5 mg, 68%), identical in all respects to authentic material: ^1H NMR (DMSO- d_6 , 400 MHz) δ 11.74 (s, 1H), 11.72 (s, 1H), 10.44 (s, 1H), 10.17 (s, 1H), 8.23 (s, 1H), 8.13 (d, $J = 8.4$ Hz, 1H), 7.99 (s, 1H), 7.87 (d, $J = 8.8$ Hz, 1H), 7.68 (d, $J = 8.0$ Hz, 1H), 7.59 (dd, $J = 8.8, 1.6$ Hz, 1H), 7.55–7.47 (m, 3H), 7.43 (s, 1H), 7.37 (t, $J = 7.6$ Hz, 1H), 7.24 (s, 1H), 7.20 (t, $J = 7.2$ Hz, 1H), 7.07 (t, $J = 7.6$ Hz, 1H), 5.28 (s, 2H), 4.83 (t, $J = 10$ Hz, 1H), 4.58 (d, $J = 7.2$ Hz, 1H), 4.24 (m, 1H), 4.04 (dd, $J = 11, 3.2$ Hz, 1H), 3.89 (dd, $J = 11, 7.2$ Hz, 1H); ^{13}C NMR (DMSO- d_6 , 125 MHz) δ 160.0, 159.5, 154.1, 142.2, 136.6, 133.3, 131.8, 131.6, 131.3, 129.8, 127.2, 127.05, 127.02, 123.4, 123.1, 123.0, 122.7, 122.1, 121.5, 119.7, 119.2, 114.9, 112.8, 112.3, 112.2, 105.6, 103.3, 100.3, 55.0, 47.6, 41.1; ESI-TOF HRMS m/z 535.1544 ($M + H^+$, $C_{31}H_{23}ClN_4O_3$ requires 535.1531).

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Supporting Information Available: General experimental details, Figure 3, and NMR spectra. This material is available free of charge via the Internet at <http://pubs.acs.org>.